

## Reduction in Functional Antibody Activity Against *Streptococcus pneumoniae* in Vaccinated Elderly Individuals Highly Correlates with Decreased IgG Antibody Avidity

Sandra Romero-Steiner, Daniel M. Musher,  
Marty S. Cetron, Lorna B. Pais, Jean E. Groover,  
Anthony E. Fiore, Brian D. Plikaytis, and  
George M. Carlone

From the Division of Bacterial and Mycotic Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia; and the Medical Service, Department of Infectious Disease, Veterans Administration Medical Center, Houston, Texas

The pneumococcal polysaccharide vaccine is recommended as a means of preventing invasive disease in the elderly. We compared responses to the 23-valent polysaccharide vaccine in 46 previously unvaccinated, healthy, institutionalized elderly persons (mean age, 85.5 years) with those in 12 healthy younger adults (mean age, 37 years) by measuring prevaccination and postvaccination serum IgG antibody concentrations (by ELISA), functional antibody activity (by opsonophagocytosis), IgG antibody avidity, and passive protection in mice. Postvaccination IgG antibody concentrations for two serotypes (6B and 19F) of the five studied (4, 6B, 14, 19F, and 23F) were significantly lower in elderly than in younger adults; however, opsonophagocytic activity was significantly reduced for all serotypes in the elderly. Sera with reduced opsonophagocytic activity (titer, <64) correlated with low IgG antibody avidity and protected mice poorly against pneumococcal challenge. In elderly persons receiving polysaccharide vaccination, there was a significant reduction in the functionality of postvaccination antibodies, and this appeared to increase with advanced age.

*Streptococcus pneumoniae* is one of the leading causes of community-acquired pneumoniae and is associated with high morbidity and mortality among the elderly [1–3]. Elderly persons are at increased risk for pneumococcal pneumonia and bacteremia [1, 4, 5]. Vaccination with the 23-valent pneumococcal polysaccharide vaccine is recommended for persons  $\geq 65$  years of age, regardless of their immunocompetence status. To date, less than one-third of the eligible elderly population has been vaccinated [4, 6]. The estimated efficacy of pneumococcal vaccine in immunocompetent elderly persons (age range, 65–75 years) is 70%–78% [7, 8], and in all elderly persons, 44%–61% [9]. Although studies have provided data about the general efficacy of the vaccine, more accurate surrogates of protection are needed to evaluate immune status in

the elderly after vaccination and to assist in developing recommendations for revaccination [4, 10].

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See editorial response by Janoff and Rubins  
on pages 289–91.

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The immune response to the polysaccharide vaccine in various age groups, including the elderly, has been studied previously [11–17]. Concentrations of antibody to pneumococcal capsular polysaccharides (PPSs) found in the elderly generally are thought to be similar to those in younger adults [12–14], although responses to certain polysaccharides may be reduced, with a more marked reduction in persons >85 years of age [14, 18]. In addition, elderly women may have lower responses than elderly men [14]. Data suggest that the protective effect of the pneumococcal vaccine diminishes with age [19]. A possible explanation is that anticapsular IgG antibodies of elderly persons may not be as effective in opsonizing pneumococci for phagocytosis. Studies of the functionality of the antibodies elicited by the 23-valent polysaccharide vaccine in older subjects [20] are very limited in number.

In the present study, titers of IgG antibody to PPSs and opsonophagocytic titers were determined prior to and following vaccination of institutionalized elderly persons and were compared with those in younger adult controls. In addition, antibody avidity and passive protection in mice were studied with use of selected sera from elderly persons who had high IgG antibody concentrations and low opsonophagocytic titers. This study contributes to the understanding of the immune response in elderly persons to the 23-valent pneumococcal

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Informed consent was obtained from all subjects. The protocols conformed to the guidelines established for human experimentation by the U.S. Department of Health and Human Services. The protocols used for animal experimentation were reviewed and approved by the Animal Use and Care Committee of the Baylor College of Medicine (Houston).

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Reprints or correspondence: George M. Carlone, Ph.D., Building 1, Room 1260, Mailstop A-36, Division of Bacterial and Mycotic Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia 30333 (GMC3@CDC.GOV).

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polysaccharide vaccine and presents opsonophagocytosis as an immune status indicator that should be included in assessment of the immune response of elderly persons to polysaccharide vaccination.

## Materials and Methods

**Study group, vaccination, and sera collection.** Forty-six elderly residents of a nursing home in Chicopee, Massachusetts, were vaccinated with the licensed 23-valent polysaccharide vaccine (Pnu-Immune 23; Lederle-Praxis-American Cyanamid, Pearl River, NY) as part of an epidemiological investigation of an outbreak of pneumonia in which parainfluenza virus and *S. pneumoniae* serotype 14 were implicated [21]. A dose of vaccine contained 25  $\mu$ g of each of 23 pneumococcal polysaccharide types.

Ages of the elderly subjects ranged from 63 to 103 years (mean, 85.5 years); 39 were female and seven were male. The study group was stratified into three age groups: group 1, 63 through 79 years ( $n = 10$ ); group 2, 80 through 89 years ( $n = 22$ ); and group 3,  $\geq 90$  years ( $n = 14$ ). Because the study population was primarily female, it was not stratified by gender. Paired sera were collected from all study participants. Sera designated as "pre" were collected within 3 days of vaccination; sera designated as "post" were collected 2–3 weeks following vaccination. Sera were stored at  $-70^{\circ}\text{C}$  until tested. All participants were healthy at the time of vaccination; none was bedridden.

**Control group.** Twelve healthy younger adults (aged 22–46 years; mean age, 37.0 years)—an equal number of men and women—were used as controls. Paired sera obtained prior to and 4 weeks following vaccination with the Pnu-Immune 23 were collected and stored at  $-70^{\circ}\text{C}$ .

**ELISA IgG antibody concentrations.** IgG antibody concentrations to PPSs from *S. pneumoniae* serotypes 4, 6B, 14, 19F, and 23F (abbreviated as PPS 4, PPS 6B, PPS 14, PPS 19F, and PPS 23F) were measured by a modified ELISA [22]. These serotypes are commonly isolated from patients with invasive disease and are associated with an increase in the frequency of resistance to drugs [3]. The 89SF standard reference serum (U.S. Food and Drug Administration, Bethesda, MD) was used to calculate serum antibody concentrations in micrograms per milliliter. Absorption of serum antibodies to the common cell-wall polysaccharide (CPS) was performed by incubation (for 30 minutes at room temperature) of diluted serum (1:50) in a solution of purified CPS (10  $\mu$ g/mL; Statens Serum Institut, Copenhagen). The substrate used was *o*-phenylenediamine dihydrochloride (Sigma, St. Louis).

**Opsonophagocytosis.** Functional antibody activity was measured in prevaccination and postvaccination sera by opsonophagocytosis (an antibody- and complement-dependent reaction), with use of differentiated HL-60 cells (granulocytes) as the effector cells [23]. HL-60 granulocytes can efficiently phagocytize and kill pneumococci, giving opsonophagocytic

titers that highly correlate with those obtained with polymorphonuclear neutrophils from donors [23]. Opsonophagocytic titers were calculated for pneumococcal serotypes 4, 6B, 14, 19F, and 23F in a viability assay as the reciprocal of the serum dilution that had  $\geq 50\%$  killing by differentiated HL-60 cells, in comparison with antibody-free complement-rich controls (12.5% per well of 3- to 4-week rabbit serum; Pel-Freez, Brown Deer, WI). All pneumococcal strains used in this study were recent clinical isolates previously used as reference strains [23].

**Antibody avidity determinations.** The relative functional antibody avidity of selected postvaccination sera from elderly and younger adults with anticapsular IgG concentrations greater than or equal to a threshold concentration of 2  $\mu$ g/mL and decreased functional antibody activity (below a threshold titer of 64) was compared with the avidity in postvaccination sera with ELISA levels of  $\geq 2$   $\mu$ g/mL and functional activity of  $\geq 64$  in opsonophagocytic titers. Antibody avidity measures the relative strength of the antigen-antibody binding. Antibody avidity can affect the measurement of ELISA IgG concentrations used in the evaluation of vaccine-induced antibodies [24, 25].

Relative antibody avidity was determined by a modification of the method previously described by MacDonald et al. [26]. In brief, Immulon IV (Dynatech, Alexandria, Va) microtiter plates were coated with 10  $\mu$ g/mL of each polysaccharide tested. A single predetermined serum concentration (from the linear portion of each serum ELISA IgG curve) was loaded onto each well in a 50- $\mu$ L volume. Subsequently, 50  $\mu$ L of a series of seven threefold dilutions of sodium thiocyanate (NaSCN; Sigma), a chaotropic compound that interferes with the antigen-antibody reaction, was added to each well, so that the final concentration ranged from 4 *M* to 0.05 *M*. Addition of NaSCN solution to the PPS-coated plates did not affect the amount of PPS bound to the plate. Plates were incubated at  $37^{\circ}\text{C}$  for 2 hours. The remainder of the assay was done following the antipneumococcal IgG-specific ELISA described above. The percentage reduction of the total absorbance (wave length, 460 nm) was calculated for each NaSCN concentration.

**Passive protection in mice.** The capacity of serum to passively protect mice against challenge with *S. pneumoniae* serogroup 4 was investigated in an adult mouse model with death as an endpoint [27]. Bacterial challenges were performed in groups of four outbred Swiss White mice (6–8 weeks old) with 10, 100, and 1,000 times the  $\text{LD}_{50}$ , 45 minutes after intraperitoneal injection of 0, 6, 18, 50, or 150 ng of IgG obtained by diluting the human sera to yield the desired dose in a final volume of 0.1 mL. One  $\text{LD}_{50}$  corresponded to 2–4 bacteria/mL. The number of surviving mice was recorded at 5 days after challenge.

**Statistical analysis.** Linear correlations were calculated with use of the Pearson's product moment correlation coefficient. Differences among groups of data were determined by

the Mann-Whitney rank-sum test, and those between pairs in 2-by-2 tables were determined by a two-tailed Fisher's exact test. Significance level was set at  $P < .05$  for all tests. The opsonophagocytic titers and ELISA IgG antibody concentrations ( $\mu\text{g/mL}$ ) were converted to a  $\log_2$  base for statistical analysis. Opsonophagocytic titers  $<8$  were reported as titers of 4 for calculation purposes. Single antibody avidity values were calculated as the weighted average of the NaSCN concentration able to reduce most of the ELISA IgG absorbance. Weights were assigned as the percent reduction of total absorbance for each serum at each NaSCN concentration. Statistical calculations were performed with use of SigmaStat software, version 1.0 (Jandel, San Rafael, CA), and Epi Info software, version 6.02 (Centers for Disease Control, Atlanta).

## Results

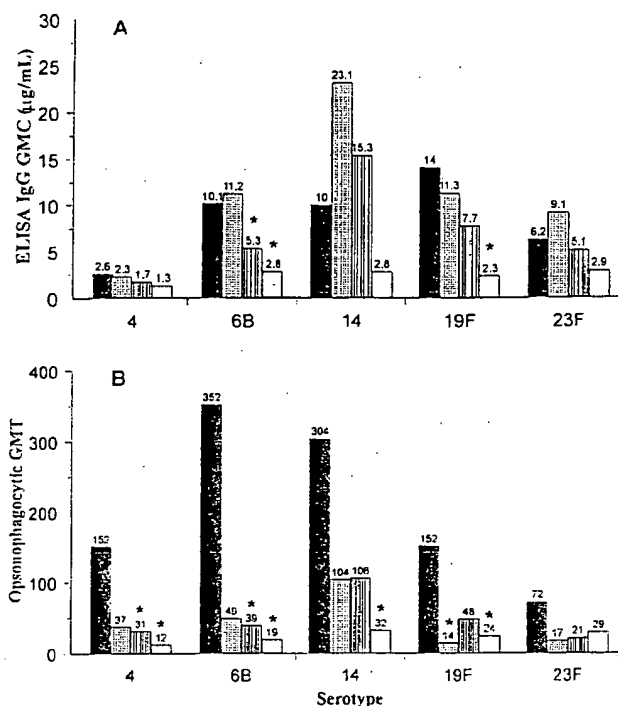
**IgG antibody detected by ELISA.** Following vaccination, elderly subjects had significant increases in concentrations of IgG antibody to all serotypes tested. Comparison of the post-vaccination IgG antibody concentrations of elderly adults with those of young adults revealed differences in the responses by age group. Table 1 gives the geometric mean concentrations (GMCs) of IgG and opsonophagocytic geometric mean titers (GMTs) for younger and elderly adults. IgG antibody GMCs in the elderly for serotypes 6B and 19F (GMCs of 5.1 and 5.8  $\mu\text{g/mL}$ , respectively) were significantly lower than those in the young adult group (10.1 and 14.0  $\mu\text{g/mL}$ , respectively).

**Table 1.** Geometric mean IgG PPS-specific serum antibody concentrations ( $\mu\text{g/mL}$ , per ELISA) and opsonophagocytic titers (reciprocal serum dilution) for elderly and young recipients of 23-valent pneumococcal polysaccharide vaccine.

Streptococcus pneumoniae serotype	Serum specimen	Value determined by indicated method, per age group			
		Young controls, aged 22–46 y (n = 12)		All elderly subjects, aged 63–103 y (n = 46)	
		ELISA	Opsono	ELISA	Opsono
4	Pre	0.8	10.4	0.8	5.2*
	Post	2.6	152.2	1.7	24.6*
6B	Pre	3.7	25.3	2.5	5.5*
	Post	10.1	352.1	5.1*	37.3*
14	Pre	0.8	19.0	2.6*	9.3*
	Post	10.0	304.4	10.0	76.6*
19F	Pre	5.2	20.1	2.9	7.1*
	Post	14.0	152.2	5.8*	28.7*
23F	Pre	2.6	7.5	2.5	7.9
	Post	6.2	71.5	4.8	22.0*

NOTE. Opsono = opsonophagocytosis; Post = postvaccination; PPS = pneumococcal capsular polysaccharide; Pre = prevaccination.

\* Significant difference ( $P < .05$ ) vs. values for the younger controls, by the Wilcoxon sample test (the Mann-Whitney rank-sum test was in agreement for the comparison between all younger adults and all elderly recipients for both ELISA IgG concentration and opsonophagocytic titer).



**Figure 1.** A, Geometric mean concentration (GMC;  $\mu\text{g/mL}$ ) of IgG pneumococcal capsular polysaccharide-specific serum antibody, as determined by ELISA, for young controls (22–46 years; n = 12; black bars) and for elderly age groups 1 (63–79 years; n = 10; dotted bars), 2 (80–89 years; n = 22; striped bars), and 3 ( $\geq 90$  years; n = 14; open bars) following vaccination with the 23-valent pneumococcal polysaccharide vaccine. B, Geometric mean opsonophagocytic titer (GMT; reciprocal serum dilution) for the young controls and elderly age groups shown in panel A, following vaccination. An asterisk designates a significant difference ( $P < .05$ ) from values for the younger controls, per the Wilcoxon sample test.

Other significant differences in IgG concentrations in elderly and young adults following vaccination included lower GMCs against serotype 6B in elderly groups 2 and 3 (5.3  $\mu\text{g/mL}$  and 2.8  $\mu\text{g/mL}$ , respectively, vs. 10.1  $\mu\text{g/mL}$  in young adults) and against serotype 19F in elderly group 3 (2.3  $\mu\text{g/mL}$ , vs. 14.0  $\mu\text{g/mL}$  in young adults) (figure 1A). Prevaccination ELISA IgG antibody concentrations were significantly different for serotype 14, when those in all elderly recipients were compared with those in younger adults.

The percentage of vaccinees attaining a twofold or higher rise in IgG antibody concentration was lower in the elderly than in young adults for serotypes 6B and 14 (table 2). However, no difference was observed in the numbers of elderly vs. young adult subjects with an IgG antibody concentration of  $\geq 2$   $\mu\text{g/mL}$  following vaccination, except for serotype 19F (9 of 14 in elderly group 3 [ $\geq 90$  years] vs. 12 of 12 young adults).

**Opsonophagocytic antibody activity.** Far more striking than the IgG antibody differences given above were the significant reductions in postvaccination opsonophagocytic GMTs for the elderly vs. young adults. For all serotypes tested, there

**Table 2.** Percentage of elderly and young recipients of 23-valent pneumococcal polysaccharide vaccine with a twofold or higher rise in ELISA IgG concentration or a fourfold or higher rise in opsonophagocytic titer between prevaccination and postvaccination sera sampling.

<i>Streptococcus pneumoniae</i> serotype	Percentage of recipients				Percentage of elderly recipients, per age group					
	Young, aged 22–46 y (n = 12)		All elderly, aged 63–103 y (n = 46)		Group 1: 63–79 y (n = 10)		Group 2: 80–89 y (n = 22)		Group 3: ≥90 y (n = 14)	
	ELISA	Opsono	ELISA	Opsono	ELISA	Opsono	ELISA	Opsono	ELISA	Opsono
4	45	73	47	42	50	60	38	48	57	21*
6B	83	83	44*	57	70	50	43*	64	29*	50
14	92	75	56*	59	50	30	67	68	43*	50
19F	75	83	41	31*	30	20*	64	38*	14*	29*
23F	50	75	33	38*	30	40	32	38	36	36

NOTE. Opsono = opsonophagocytosis.

\* Significant difference ( $P < .05$ ) between young controls and elderly groups, per Fisher's exact two-tailed test. Prevaccination IgG concentrations ( $\mu\text{g/mL}$ ) in the elderly were significantly elevated only for serotype 14.

was a significant reduction in opsonophagocytic GMTs for the elderly ( $P < .05$ ) when compared with GMTs for young adults (table 1). Differences in opsonophagocytic GMTs were more evident as age increased: in group 1, differences were significant only for serotype 19F; in group 2, for serotypes 4 and 6B; and in group 3, for all serotypes except 23F (figure 1B).

Prevaccination opsonophagocytic titers of all elderly recipients were also significantly lower for all serotypes, except serotype 23F, when compared with those for younger adults (table 1). A significantly lower percentage of elderly vaccine recipients than young controls attained a fourfold or higher rise in opsonophagocytic titers (after vaccination) for serotypes 4, 19F, and 23F (table 2).

Serum IgG antibody concentrations ( $\log_2$ ) in elderly and young adults had significant ( $P < .05$ ) correlation coefficients of association ( $r$  values) with opsonophagocytic titers ( $\log_2$ ) for all the serotypes tested (figure 2). However, post-vaccination  $r$  values for serotypes 4, 6B, 19F, and 23F were lower for sera from elderly persons ( $r$  values ranged from .30 to .62) than for sera from young adults ( $r$  values ranged from .61 to .86). This reduction in correlation coefficients was due to a higher number of serum specimens from elderly persons who had IgG antibody concentrations of  $\geq 2 \mu\text{g/mL}$  and opsonophagocytic titers of  $< 64$ . The proportion of sera with opsonophagocytic titers of  $\geq 64$ , as well as IgG antibody concentrations of  $\geq 2 \mu\text{g/mL}$ , was significantly reduced in most elderly groups for serotype 6B (21%–45% in the elderly groups vs. 92% in young adults), serotype 19F (30%–42% for elderly groups vs. 83% in young adults), and serotype 14 (36% in elderly group 3 vs. 83% in young adults). To determine reasons for the differences in functional antibody activity observed between elderly and young adults, we further analyzed these sera to assess antibody avidity measurements and passive protection in mice.

**Antibody avidity measurements.** For the avidity assays performed in sera from elderly persons with low opsonophagocytic activity, low concentrations of NaSCN (range,

0.005  $M$ –0.15  $M$ ) were able to reduce ELISA optical density by 85% (SD,  $\pm 13.2\%$ ). This suggested that the antibody measured by ELISA was of low avidity. In 37 serum assays that showed a discrepancy between IgG antibody concentration and opsonophagocytic titer, 34 (92%) of the specimens had low antibody avidity. In contrast, in sera with anticapsular antibody of  $\geq 2 \mu\text{g/mL}$  (threshold IgG concentration) and a high opsonophagocytic titer ( $\geq 64$  threshold titer), the reaction between IgG antibody and type-specific PPS was significantly inhibited only by higher NaSCN concentrations ( $\geq 0.44 M$ ), suggesting the presence of high-avidity antibody in the sera. Figure 3A shows a significant correlation ( $r = .76$ ,  $P < .01$ ) between opsonophagocytic titer ( $\log_2$ ) and the weighted average of NaSCN concentration. There was a significant correlation between ELISA IgG antibody concentration and the weighted average of NaSCN concentrations ( $r = .36$ ,  $P < .01$ ), although the  $r$  value was lower than that obtained with opsonophagocytosis (figure 3B). The association of low antibody avidity and low opsonophagocytic antibody activity was observed in sera from elderly and young adults.

**Passive protection in mice.** A high opsonophagocytic titer and high avidity for capsular PPS antigens were found to confer protection against pneumococcal challenge in mice. Thus, for example, serum 7023 (a specimen from an elderly subject that had high avidity and high opsonophagocytic activity) protected mice at all doses tested (table 3). In contrast, identical doses of IgG antibody from serum 7047, a specimen with low avidity and low opsonophagocytic activity from an elderly person, failed to protect even at the lowest inoculum of pneumococci. Sera with IgG antibody of intermediate avidity exhibited intermediate degrees of protection (data not shown). Because all sera tested had similar IgG antibody concentrations and yet conferred varying degrees of protection in mice, no association was observed between ELISA IgG antibody concentration and protection against death.

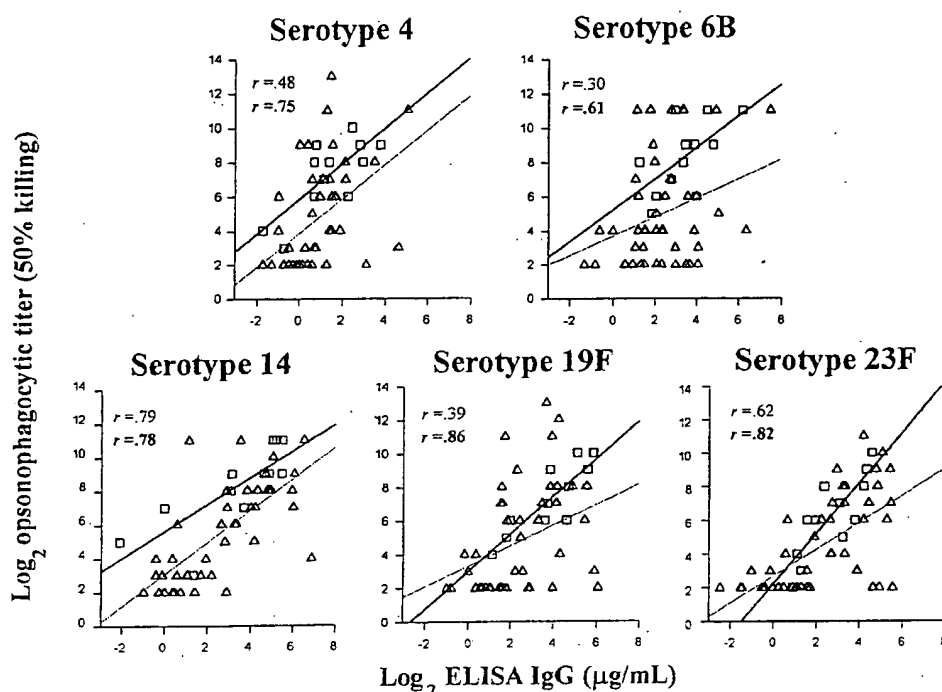


Figure 2. Correlation between  $\log_2$  ELISA IgG antibody concentration ( $\mu\text{g/mL}$ ) and  $\log_2$  opsonophagocytic titer in postvaccination sera from young controls ( $n = 12$ ; squares) and elderly recipients of vaccine against *Streptococcus pneumoniae* serotypes 4, 6B, 14, 19F, and 23F ( $n = 46$ ; triangles). Linear regression for sera from young adults is indicated by the bold line, whereas the other line designates the linear regression for sera from elderly adults.

## Discussion

Our observations indicate that IgG antibody to PPSs is elicited in most elderly persons following vaccination. Similar percentages of elderly and young adults had IgG antibody at or above a threshold concentration of  $2 \mu\text{g/mL}$  to four of the five PPSs tested. However, the IgG antibody GMCs were lower for two PPSs, and the ability to elicit twofold or higher increases in IgG antibody among the elderly was lower for two PPSs. This suggests that although responsiveness to polysaccharide antigens is retained with increasing age, it may be at a diminished level.

It is possible that the reduction observed in the elderly was due to the measurement of antibodies at 2–3 weeks instead of 4 weeks postvaccination. However, Musher et al. had previously shown only small differences in the IgG antibody concentration measured at 14 vs. 27 days postvaccination [12]. Earlier studies showed less [14] or no [12, 28] diminution in the antibody response with aging; differences in our results may reflect the fact that most of our elderly participants were  $>80$  years of age. In addition, the predominance of women in our study group may have been a factor; Sankilampi et al. [14] found that women have slightly lower antibody responses to some pneumococcal polysaccharide antigens.

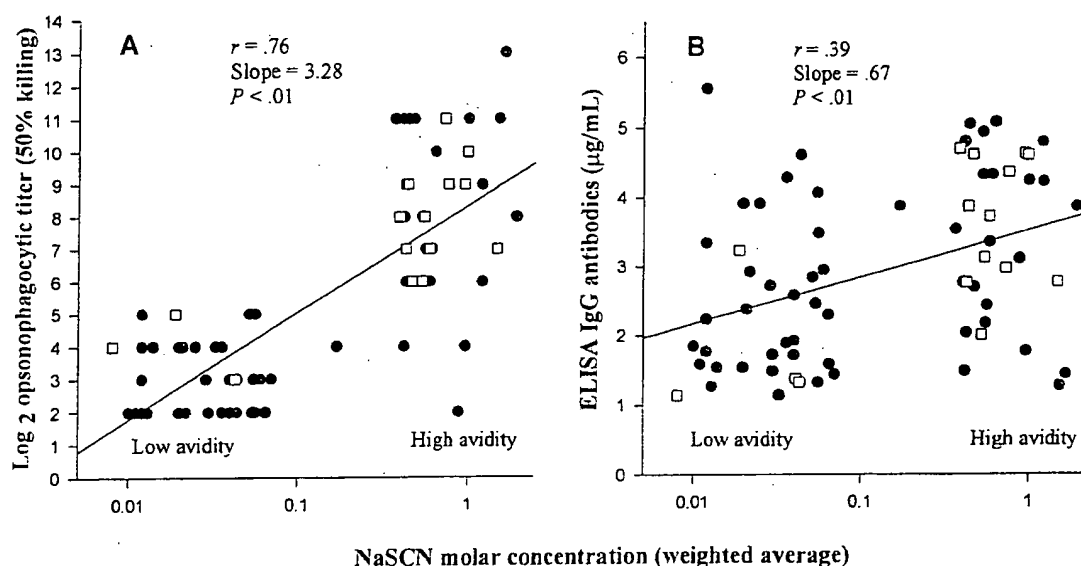
A more important finding of the present study was that the ability to elicit a functional antibody response is distinctly

reduced with advanced age, as was shown by significantly lower opsonophagocytic GMCs in the elderly than in young adults to all the serotypes tested. This type of reduction was noticed in only  $\sim 20\%$  of the elderly studied in a recent investigation by Rubins et al. [28]; however, only serotype 14 was assessed for functional antibody activity.

Our observations indicate that functional opsonophagocytic activity to serotype 14 highly correlates with ELISA IgG antibody concentrations (figure 2) and that PPS 14 elicits antibodies of higher avidity in both elderly and younger adults than do other PPSs commonly tested. Therefore, significant differences in functionality (opsonophagocytosis and/or antibody avidity) of the antibodies generated in the elderly may not be observed by studying only this serotype.

Our study suggests that the ability to elicit a fourfold or higher rise in opsonophagocytic antibody titers following vaccination decreased with age for three of the five serotypes tested. In addition, the number of elderly persons with an opsonophagocytic titer  $\geq 64$  and an ELISA IgG antibody concentration  $\geq 2 \mu\text{g/mL}$  was lower for serotypes 6B and 19F, as well as for serotype 14 in the very elderly. For serotypes 4 and 23F the percentage of elderly recipients with high opsonophagocytic titers decreased, although not significantly (data not shown).

These observations were strengthened by the results of the



**Figure 3.** A. Correlation between the weighted average of the sodium thiocyanate (NaSCN) molar concentration yielding most of the reduction in ELISA IgG absorbance and the log<sub>2</sub> opsonophagocytic titer. Results represent all serotypes (combined) in selected sera from elderly subjects (black circles) and young adults (squares). The line designates the linear regression. All sera with high antibody avidity also had high opsonophagocytic titers ( $\geq 64$ ), except for sera from three elderly subjects that were tested against *Streptococcus pneumoniae* serogroup 4 and had low opsonophagocytic titers ( $< 64$ ) and high antibody avidity (weighted averages: 0.42 M, 0.88 M, and 0.96 M NaSCN). All serum samples shown had ELISA IgG antibody concentrations  $\geq 2$  µg/mL. B. Correlation between the weighted average of the NaSCN molar concentrations yielding most of the reduction in ELISA IgG absorbance and the ELISA IgG antibody concentration (µg/mL). Symbols are the same as in panel A. Sera with low antibody avidity (0.01 M to 0.1 M NaSCN) also had low opsonophagocytic titers ( $< 64$ ), regardless of the ELISA IgG antibody concentration. Sera with high antibody avidity ( $> 0.4$  M NaSCN) also had high opsonophagocytic titers ( $\geq 64$ ).

passive-protection experiments in mice with use of *S. pneumoniae* serotype 4, in which 100% protection was achieved by administering a serum with high opsonophagocytic titer and antibody avidity; no protection was observed with a serum that had low opsonophagocytic titer and antibody avidity. Similar patterns of passive protection were observed for serotype 6B with sera from elderly and young adults in an infant mouse model of bacteremia (data not shown).

Differences between IgG antibody measured by ELISA and functional opsonophagocytic activity in sera from elderly persons appeared to be related to differences in the avidity of IgG for PPS. Low concentrations of NaSCN readily inhibited the interaction between PPSs and IgG antibody from sera with low opsonophagocytic activity, whereas much higher concentrations of NaSCN were needed to produce such inhibition in sera with high opsonophagocytic activity, such as antibodies against serotype 14.

**Table 3.** Passive protection capacity of sera from elderly persons with known ELISA IgG antibody concentration, opsonophagocytic titer, and antibody avidity against *Streptococcus pneumoniae* serotype 4.

Serum no.	Age (y) of patient	Dose of IgG antibody to serotype 4 (ng)	Percentage of mice surviving* after challenge with			Opsonic titer (50% killing)	ELISA IgG (µg/mL)	Antibody avidity: NaSCN <sup>†</sup>
			10 × LD <sub>50</sub>	100 × LD <sub>50</sub>	1,000 × LD <sub>50</sub>			
7023	90	150	100 (4/4)	100 (4/4)	100 (4/4)	8,192	2.7	1.67
		50	100 (4/4)	100 (4/4)	100 (4/4)			
		18	100 (4/4)	100 (4/4)	100 (4/4)			
		6	100 (4/4)	100 (4/4)	100 (4/4)			
7047	74	150	25 (1/4)	0 (0/4)	0 (0/4)	4	2.4	0.013
		50	50 (2/4)	0 (0/4)	0 (0/4)			
		18	50 (2/4)	0 (0/4)	0 (0/4)			
		6	0 (0/4)	0 (0/4)	0 (0/4)			

NOTE. LD<sub>50</sub> = lethal dose for 50% of adult mice (1 LD<sub>50</sub> = 2–4 bacteria/mL).

\* In parentheses is the no. of surviving mice/total no. of mice challenged.

<sup>†</sup> Molar concentration of sodium thiocyanate (NaSCN) necessary to yield an 85% reduction in ELISA IgG optical density.

In a study of older adults, Konradsen found no notable discrepancies between antibody concentration and avidity for various PPSs [29]; however, this finding may reflect the use of subjects 60–67 years of age, 20–40 years younger than the elderly subjects in our study. Similarly, Rubins et al. found no difference between elderly and younger adults in terms of serum antibody concentration and antibody avidity to serotype 14 [28]; however, as mentioned above, this serotype rarely elicits antibodies of low avidity in the elderly. Our study indicates that only nine of 46 elderly recipients had an opsonophagocytic titer  $<64$  and an ELISA IgG antibody concentration  $\geq 2 \mu\text{g/mL}$  against PPS 14. Neither one of these studies made a direct correlation between opsonophagocytic activity and antibody avidity.

In the immunogenicity studies reported to date, the elderly have responded equally to conjugated and unconjugated polysaccharides [30, 31]; however, the functional antibody response has not been addressed in these studies. If the poor functionality of antibodies to PPSs as humans age is caused by an adherent accessory cell deficiency for antigenic presentation in the spleen, as has been shown in mice [32], a vaccination strategy may not be able to overcome the defect. Reduced neutrophil function in the elderly (reduced activation of superoxide anions and increased level of apoptosis) could account for the increased risk for pneumococcal infections, regardless of vaccination status [33, 34].

In summary, this study highlights the importance of evaluating the IgG antibody response, as well as the functional antibody activity of the antibodies measured, especially in high-risk populations. The distinct reduction in functional IgG antibody activity in the elderly was more pronounced in those 80–89 years of age and  $\geq 90$  years of age. However, these age groups represent only 21% and 4.3%, respectively, of the target U.S. elderly population (32.4 million persons  $\geq 65$  years old) for the pneumococcal polysaccharide vaccine [35].

Thus, this study should not discourage the use of the 23-valent polysaccharide vaccine in the elderly until a better vaccination strategy is available. New and improved approaches to vaccination in the elderly should be considered.

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#### References

- Fein AM. Pneumonia in the elderly: special diagnostic and therapeutic considerations. *Med Clin North Am* 1994;78:1015–33.
- Marrie TJ. New aspects of old pathogens of pneumonia. *Med Clin North Am* 1994;78:987–95.
- Breiman RF, Butler JC, Tenover FC, Elliott JA, Facklam RR. Emergence of drug-resistant pneumococcal infections in the United States. *JAMA* 1994;271:1831–5.
- Centers for Disease Control and Prevention. Prevention of pneumococcal disease: recommendations of the Advisory Committee on Immunization Practices (ACIP). *MMWR Morb Mortal Wkly Rep* 1997;46(RR-8):1–24.
- Centers for Disease Control and Prevention. Defining the public health impact of drug-resistant *Streptococcus pneumoniae*: report of a working group. *MMWR Morb Mortal Wkly Rep* 1996;45(RR-1):1–20.
- Centers for Disease Control and Prevention. Pneumococcal and influenza vaccination levels among adults aged  $\geq 65$  years—United States, 1995. *MMWR Morb Mortal Wkly Rep* 1997;46(39):913–9.
- Sims RV, Steinmann WC, McConville JH, King LR, Zwick WC, Schwartz JS. The clinical effectiveness of polyvalent pneumococcal vaccine in the elderly. *Ann Intern Med* 1988;108:653–7.
- Butler JC, Breiman RF, Campbell JF, Lipman HB, Broome CV, Facklam RR. Pneumococcal polysaccharide vaccine efficacy: an evaluation of current recommendations. *JAMA* 1993;270:1826–31.
- Stein BE. Vaccinating elderly people: protecting from avoidable disease. *Drugs Aging* 1994;5:242–53.
- Wenger JD, Steiner SR, Pais LB, et al. Laboratory correlates for protective efficacy of pneumococcal vaccines: how can they be identified and validated? [abstract G37]. In: Program and abstracts of the 36th Interscience Conference on Antimicrobial Agents and Chemotherapy. Washington, DC: American Society for Microbiology, 1996:150.
- Musher DM, Luchi MJ, Watson DA, Hamilton R, Baughn RE. Pneumococcal polysaccharide vaccine in young adults and older bronchitics: determination of IgG responses by ELISA and the effect of adsorption of serum with non-type-specific cell wall polysaccharide. *J Infect Dis* 1990;161:728–35.
- Musher DM, Groover JE, Rowland JM, et al. Antibody to capsular polysaccharides of *Streptococcus pneumoniae*: prevalence, persistence, and response to revaccination. *Clin Infect Dis* 1993;17:66–73.
- Ruben FL, Uhrin M. Specific immunoglobulin-class antibody responses in the elderly before and after 14-valent pneumococcal vaccine. *J Infect Dis* 1985;151:845–9.
- Sankilampi U, Honkanen PO, Bloigu A, Herva E, Leinonen M. Antibody response to pneumococcal capsular polysaccharide vaccine in the elderly. *J Infect Dis* 1996;173:387–93.
- Mufson MA, Krause HE, Schiffman G. Long-term persistence of antibody following immunization with pneumococcal polysaccharide vaccine. *Proc Soc Exp Biol Med* 1983;173:270–5.
- Roghmann KJ, Tabloski PA, Bently DW, Schiffman G. Immune response of elderly adults to pneumococcus: variation by age, sex, and functional impairment. *J Gerontol* 1987;42:265–70.
- Landesman SH, Schiffman G. Assessment of the antibody response to pneumococcal vaccine in high-risk populations. *Rev Infect Dis* 1981;3(suppl):S184–96.
- Hedlund JU, Mats EK, Örtqvist AB, Henrichsen J. Antibody response to pneumococcal vaccine in middle-aged and elderly patients recently treated for pneumonia. *Arch Intern Med* 1994;154:1961–5.
- Shapiro ED, Berg AT, Austrian R, et al. The protective efficacy of polyvalent pneumococcal polysaccharide vaccine. *N Engl J Med* 1991;325:1453–60.
- Musher DM, Chapman AJ, Goree A, Jonsson S, Briles D, Baughn RE. Natural and vaccine-related immunity to *Streptococcus pneumoniae*. *J Infect Dis* 1986;154:245–56.
- Fiore AE, Iverson C, Messmer T, et al. Outbreak of pneumonia in a long-term care facility: antecedent human parainfluenza virus 1 infection may predispose to bacterial pneumonia. *J Am Geriatr Soc* 1998;46:1112–7.
- Quartaert SA, Kirch CS, Quackenbush Weidl LJ, et al. Assignment of weight-based antibody units to a human antipneumococcal standard reference serum, lot 89-S. *Clin Diagn Lab Immunol* 1995;2:590–7.
- Romero-Steiner S, LiButti D, Pais LB, et al. Standardization of an opsonophagocytic assay for the measurement of functional antibody activity against *Streptococcus pneumoniae* using differentiated HL-60 cells. *Clin Diagn Lab Immunol* 1997;4:415–22.

24. Anttila M, Eskola J, Ahman H, Käyhty H. Avidity of IgG for *Streptococcus pneumoniae* type 6B and 23F polysaccharides in infants primed with pneumococcal conjugates and boosted with polysaccharide or conjugate vaccines. *J Infect Dis* 1998;177:1614-21.
25. Granoff DM, Maslanka SE, Carlone GM, et al. A modified enzyme-linked immunosorbent assay for measurement of antibody responses to meningococcal C polysaccharide that correlate with bactericidal responses. *Clin Diagn Lab Immunol* 1998;5:479-85.
26. MacDonald RA, Hosking CS, Jones CL. The measurement of relative antibody affinity by ELISA using thiocyanate elution. *J Immunol Methods* 1988;106:191-4.
27. Musher DM, Johnson B Jr, Watson DA. Quantitative relationship between anticapsular antibody measured by enzyme-linked immunosorbent assay or radioimmunoassay and protection of mice against challenge with *Streptococcus pneumoniae* serotype 4. *Infect Immun* 1990;58:3871-6.
28. Rubins JB, Puri AKG, Loch J, et al. Magnitude, duration, quality, and function of pneumococcal vaccine responses in elderly adults. *J Infect Dis* 1998;178:431-40.
29. Konradsen HB. Quantity and avidity of pneumococcal antibodies before and up to 5 years after pneumococcal vaccination of elderly persons. *Clin Infect Dis* 1995;21:616-20.
30. Powers DC, Anderson EL, Lottenbach K, Mink CAM. Reactogenicity and immunogenicity of a protein-conjugated pneumococcal oligosaccharide vaccine in older adults. *J Infect Dis* 1996;173:1014-8.
31. Shelly MA, Jacoby H, Riley GJ, Graves BT, Pichichero M, Treanor JJ. Comparison of pneumococcal polysaccharide and CRM<sub>197</sub>-conjugated pneumococcal oligosaccharide vaccines in young and elderly adults. *Infect Immun* 1997;65:242-7.
32. Garg M, Luo W, Kaplan AM, Bondada S. Cellular basis of decreased immune-responses to pneumococcal vaccines in aged mice. *Infect Immun* 1996;64:4456-62.
33. Ito Y, Ponnappan U, Lipschitz DA. Excess formation of lysophosphatidic acid with age inhibits myristic acid-induced superoxide anion generation in intact human neutrophils. *FEBS Lett* 1996;394:149-52.
34. Fulop T Jr, Fouquet C, Allaire P, et al. Changes in apoptosis of human polymorphonuclear granulocytes with aging. *Mech Aging Dev* 1997;96:15-34.
35. Bureau for the Census. Population estimates/nation/intfile2-1.txt. Worldwide web site <http://www.census.gov>. Accessed 14 April 1998.